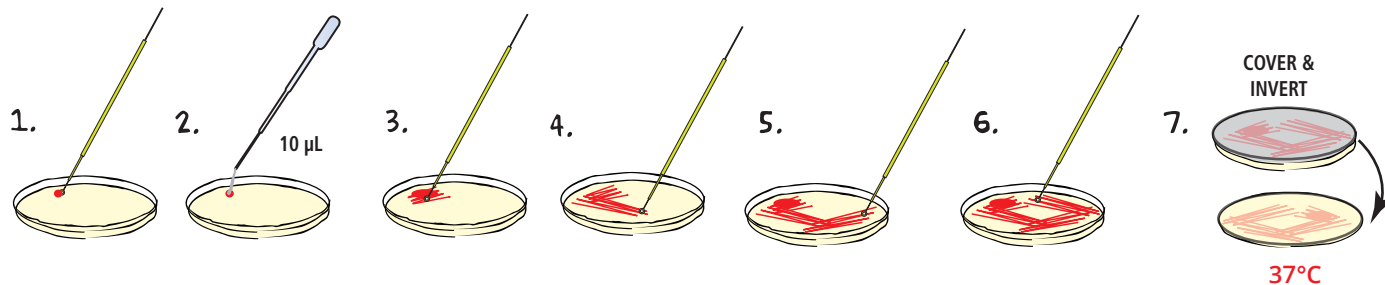


# INSTRUCTION GUIDE: Using BactoBeads™

*BactoBeads™ are non-pathogenic, lyophilized bacteria that can easily be cultured for use in the classroom.*

**IMPORTANT** – BactoBeads™ absorb moisture if left open, even for a short period of time. Open the vial only when you are ready to use the beads. If you are not using all of the beads at one session, quickly remove the required number of beads and promptly/tightly secure the gray stopper back to the vial. Place the unused vial of beads in a Ziploc bag with desiccant and store at room temperature. If noticeable moisture has caused the beads to shrink in size, they may not function properly.



1. **OPEN** the vial of Bactobead™ by carefully removing the metal crimp the gray rubber stopper. **REMOVE** a single BactoBead™ from the vial using a sterile inoculating loop. Using aseptic technique, **TRANSFER** the bead to the edge of a large petri plate (LB source plate) and replace lid. **CAP** the vial immediately after using to limit exposure to moisture in the air.
2. Instantly **DISSOLVE** the bead by adding 10 µL of sterile liquid broth or sterile water.
3. **STREAK** the loop back and forth through the dissolved BactoBead™ to make a primary streak at the top of the plate. Try not to gouge the loop into the medium.
4. **STREAK** the loop through primary streak to a clean part of the agar several times to create a secondary streak.
5. **ROTATE** the plate. **STREAK** the loop through the secondary streak to a clean part of the agar several times.
6. **ROTATE** the plate once more. **STREAK** the loop through the third streak to a clean part of the agar. This should produce isolated colonies.
7. **COVER** the plate and **INCUBATE INVERTED** at 37 °C for 16 to 18 hours. If you do not have an incubator, colonies will form at room temperature in approximately 24 - 48 hours.
8. **REPEAT** the above steps for the number of plates needed. Alternatively, use a sterile swab to inoculate the entire plate (if isolated colonies are not needed).

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